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## Protozoan, Helminth, and Arthropod Parasites of the Spotted Chorus Frog, *Pseudacris clarkii* (Anura: Hylidae), from North-central Texas

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ABSTRACT: Thirty-nine juvenile and adult spotted chorus frogs, *Pseudacris clarkii*, were collected from 3 counties of north-central Texas and examined for parasites. Thirty-three (85%) of the *P. clarkii* were found to be infected with 1 or more parasites, including *Hexamita intestinalis* Dujardin, 1841, *Tritrichomonas augusta* Alexeieff, 1911, *Opalina* sp. Purkinje and Valentin, 1840, *Nyctotherus cordiformis* Ehrenberg, 1838, *Myxidium serotinum* Kudo and Sprague, 1940, *Cylindrotaenia americana* Jewell, 1916, *Cosmocercoides variabilis* (Harwood, 1930) Travassos, 1931, and *Hannemania* sp. Oudemans, 1911. All represent new host records for the respective parasites. In addition, a summary of the 36 species of amphibians and reptiles reported to be hosts of *Cylindrotaenia americana* is presented.

KEY WORDS: Anura, Cosmocercoides variabilis, Cylindrotaenia americana, Hannemania sp., Hexamita intestinalis, Hylidae, intensity, Myxidium serotinum, Nyctotherus cordiformis, Opalina sp., prevalence, Pseudacris clarkii, spotted chorus frog, survey, Tritrichomonas augusta.

The spotted chorus frog, Pseudacris clarkii (Baird, 1854), is a small, secretive, hylid anuran that ranges from north-central Kansas southward through central Oklahoma and Texas to northeastern Tamaulipas, Mexico (Conant, 1975). The species inhabits marshy areas of open prairie grasslands and edges of woodland. Little information is available on the ecology and natural history of P. clarkii (Pierce and Whitehurst, 1990), and even less is known about its parasites. Kuntz and Self (1944) examined 3 P. clarkii from Comanche County, Oklahoma, for helminth parasites and reported Glypthelmins quieta (Stafford, 1900) Stafford, 1905, in a single frog. To my knowledge, nothing else has been published regarding either ecto- or endoparasites of this frog. The purposes of this paper are to report the identity, prevalence, and intensity of parasites infecting P. clarkii in north-central Texas, and to provide a summary of the amphibians and reptiles of the world known to be hosts of Cylindrotaenia americana.

#### **Materials and Methods**

Between May 1986 and April 1988, and again during March 1990, 39 juvenile and adult *P. clarkii* (24 males, 15 females;  $\bar{x} \pm$  SE snout-vent length [SVL] = 28.7  $\pm$ 0.4, range = 24-33 mm) were collected from Dallas (*N* = 14), Hood (*N* = 12), and Somervell (*N* = 13) counties of north-central Texas and examined for parasites. Specimens were either taken by hand by overturning limestone rocks or were captured with dipnets in temporary ponds during spring breeding activities. Frogs were placed in plastic freezer bags on ice and trans-

ported to the laboratory where they were killed with an overdose of Nembutal<sup>®</sup>. Necropsy and parasite techniques are identical to the methods of McAllister (1987) and McAllister and Upton (1987a, b), except that cestodes were stained with Semichon's acetocarmine and larval chiggers were fixed in situ with 10% formalin, sectioned at 7  $\mu$ m, and stained with hematoxylin and eosin counterstain. Voucher specimens of frogs have been deposited in the Arkansas State University Museum of Zoology (ASUMZ 5948, 5966, 5969, 5977-5979, 6015-6016, 7069-7072, 8515, 8578, 8628-8634, 8637, 8683-8688, 8691). Parasites have been deposited in the U.S. National Parasite Collection, USDA, Beltsville, Maryland 20705 as follows: Hexamita intestinalis (USNM 81062), Tritrichomonas augusta (USNM 81063), Opalina sp. (USNM 81059), Nyctotherus cordiformis (USNM 81060), Myxidium serotinum (USNM 81061), Cylindrotaenia americana (USNM 81057-81058), Cosmocercoides variabilis (USNM 81065), Hannemania sp. (USNM 81064).

#### **Results and Discussion**

Thirty-three (85%) of the *P. clarkii* harbored 1 or more parasites (Table 1); all are new host records. None of the *P. clarkii* was found to be infected with coccidial parasites in the feces and the blood was negative for intraerythrocytic or trypanosomal hematozoa. Of frogs from the 3 separate county locations, 14 (100%) from Dallas, 11 (92%) from Hood, and 8 (62%) from Somervell counties were infected.

The cosmopolitan flagellates, *Hexamita intestinalis* Dujardin, 1841, and *Tritrichomonas augusta* Alexeieff, 1911, were the most common parasites of *P. clarkii*. Both species have been reported previously from a number of amphib-

Parasite	Site of infection*	Prevalence <sup>†</sup>
Protozoa		
Mastigophora		
Hexamita intestinalis	CO, RE	33/39 (85%)
Tritrichomonas augusta	CO, RE	33/39 (85%)
Opalinata		
Opalina sp.	CO	17/39 (44%)
Ciliophora		
Nyctotherus cordiformis	CO	2/39 (5%)
Мухоzоа		
Myxidium serotinum	GB	23/39 (59%)
Platyhelminthes		
Cestoidea		
Cylindrotaenia americana	SI	3/39 (8%)
Nematoda		
Cosmocercoides variabilis	RE	2/39 (5%)
Acari		
Hannemania sp. (larvae)	DE	11/39 (28%)

Table 1. Parasites found in Pseudacris clarkii from north-central Texas.

\* Abbreviations: CO, colon; DE, dermis; GB, gall bladder; RE, rectum; SI, small intestine.

<sup>†</sup> Number infected/number examined (Margolis et al., 1982).

ians (Buttrey, 1954; Frank, 1984), including Brimley's chorus frogs, *P. brimleyi*, from North Carolina (Brandt, 1936) and western chorus frogs, *P. triseriata triseriata*, from Ohio (Odlaug, 1954).

The endocommensal *Opalina* sp. Purkinje and Valentin, 1840, was found in nearly half of the frogs examined. McAllister (1987) reported that 51 of 52 (98%) Strecker's chorus frogs, *P. streckeri streckeri*, from Dallas County, Texas, were infected with *Opalina* sp. Interestingly, the opalinids noted herein were morphologically indistinguishable from those of sympatric *P. s. streckeri*. However, as noted by McAllister (1987), specific identification was not possible.

Only 5% of the *P. clarkii* were found to be infected with *Nyctotherus cordiformis* Ehrenberg, 1838. However, a 10-fold higher prevalence of *N. cordiformis* was reported for *P. s. streckeri* (McAllister, 1987), as 54% were infected. This ciliate has been reported from other *Pseudacris* spp. (Walton, 1964).

Spores and trophozoites of *Myxidium serotinum* Kudo and Sprague, 1940, were found in more than half of the frogs. A moderately high prevalence of this myxozoan was also reported for *P. s. streckeri* (McAllister, 1987). Additional amphibians from north-central Texas have been reported previously to harbor *M. serotinum*, including smallmouth salamanders, *Ambystoma texanum* (McAllister and Upton, 1987b), and 3 species of toads, *Bufo* spp. (McAllister et al., 1989).

Adult nematotaeniid tapeworms, Cylindrotaenia americana Jewell, 1916, were found in 3 frogs, 1 each collected on 16 March and 10 April 1987 and on 22 June 1986 from Dallas (30 mm SVL male, ASUMZ 8684), Hood (25 mm SVL male, ASUMZ 7071), and Somervell (30 mm SVL male, ASUMZ 6016) counties, respectively. The mean intensity was 2.7 (range = 1-6) worms per host. This cestode has been reported previously from Gastrophryne olivacea (McAllister and Upton, 1987b) and Ambystoma texanum (Mc-Allister and Upton, 1987a) from north-central Texas. None of the P. s. streckeri examined by McAllister (1987) was found to be infected with C. americana, although sympatric salamanders and other frogs harbored the parasite. Cylindrotaenia americana has been reported from other amphibians and 2 reptiles from North and South America, Europe, and Asia, including 9 species of salamanders, 25 species of frogs and toads, 1 skink, and 1 snake (Table 2).

Joyeux (1924) reported C. americana from the puddle frog, Phrynobatrachus (syn. Arthroleptis) ogoensis Boulenger, 1906, and mascarene grass frog, Ptychadena mascareniensis (syn. Rana aequiplicata) Werner, 1898, from Mozambique, Africa. Harwood (1932) questioned the identity of Joyeux's material and Joyeux (in Baer, 1933)

# Table 2. Amphibians and reptiles of the world reported to be hosts of Cylindrotaenia americana Jewell, 1916.

Host	Locality	Reference
Amphibia		
Caudata		
Ambystomatidae		
Ambystoma texanum	Texas	McAllister and Upton, 1987b
Plethodontidae		
Desmognathus fuscus	North Carolina	Mann, 1932
Demonstrate	New York	Fischthal, 1955
D. monticola	I ennessee	Dunbar and Moore, 1979
D. augdramanulata	North Carolina	Goater et al., 1987
D. quaaramaculala	Tennessee	Goater et al., 1987
D. Ochrophaeus	North Carolina	Conten at al 1087
Plethodon cinercus	Tennessee	Dunber and Moore 1070
P olutinosus	Tennessee	Dunbar and Moore, 1979
P jordani	North Carolina	Dundar and Moore, 1979
P richmondi	Tennessee	Dupper and Moore 1070
Anura	Tennessee	Duiloar and Moore, 1979
Pelobatidae		
Scaphionus multiplicatus	Mexico	Walton 1940
Lentodactylidae	MOADO	Wallon, 1740
Lentodactylus ocellatus	Argentina Brazil	Savazzini 1929
Lepionaciyus Ocenaus	Argentina, Diazit	Savalliii, 1727
Hylidae		
Acris crepitans	Illinois, Michigan	Jewell, 1916
	Nebraska	Brooks, 1976a
	Iowa	Ulmer and James, 1976
	Texas	McAllister and Upton, 1987a
A. gryllus	Texas	Harwood, 1932
	Oklahoma	Trowbridge and Hefley, 1934
Hyla arborea	Czechoslovakia	Prokopic, 1957
H. arenicolor	Utah	Parry and Grundmann, 1965
H. squirella	Texas	Harwood, 1932
Pseudacris clarkii	Texas	McAllister, this study
P. triseriata	Texas	Harwood, 1932
Bufonidae		
Bufo americanus	Iowa	Ulmer and James, 1976
B. canorus	California	Ingles, 1936; Walton, 1941
B. compactilis	Mexico	Walton, 1940
B. ictericus	Brazil	Stumpf, 1981/1982a, 1981/1982b
B. marinus	Colombia	Brooks, 1976b
	Ecuador	Dyer, 1986
B. microscaphus	Utah	Parry and Grundmann, 1965
B. terrestris	SE United States	Jewell, 1916
B. typhonius	Ecuador	Dyer, 1986
B. woodhousii	Virginia	Campbell, 1968
Melanophryniscus stelzneri	Uruguay	Mane-Garzon and Gonzalez, 1978
Ranidae		• • • • • • • •
Rana aurora	Oregon, Washington	Lehmann, 1965
R. catesbeiana	Massachusetts	Rankin, 1945
<b>D</b>	Virginia	Campbell, 1968
K. pipiens	Illinois, Michigan, Nebraska	Jewell, 1916
	Oregon, washington	Lenmann, 1965
P protiona	Iowa	Unter and James, 1976
R. preuosa R. sontentrionalis	Oregon, wasnington	Lenmann, 1965 Roughard, 1951
K. septentrionalis	maine	Douchard, 1951
Gastrophrune olivesse	Texas	McAllister and Linton 1007-
Rentilia	10,43	Michanister and Opton, 198/a
Sauria		
Saulia		
Scincella lateralis	Texas	Harwood 1932
Scincena iateralis	Florida	Brooks 1072
Serpentes	TIONIda	DIOURS, 1772
Colubridae		
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synonymized it with Barietta jaegerskioeldi (Janicki, 1928) Hsü, 1935. However, Mettrick (1953) assigned Joyeux's specimens to B. janicki (Hilmy, 1936) Douglas, 1958 (see also Fischthal and Asres, 1970). Nevertheless, Yamaguti (1959) recognizes them as B. jaegerskioeldi, while Schmidt (1986) lists Arthroleptis as a host of C. americana. Dyer (1986) was apparently unaware of this confusion although he did suggest that the identity of the Joyeux's African form of C. americana with the American form seemed unlikely. In view of this controversy over Joyeux's material, neither of the above hosts is listed in Table 2.

Two nematodes, Cosmocercoides variabilis (Harwood, 1930) Travassos, 1931, were each found in 2 P. clarkii (26 mm SVL male, ASUMZ 8691; 28 mm SVL female, uncatalogued) collected on 7 March 1987 and 18 March 1990 in Somervell and Hood counties, respectively. Pseudacris clarkii could represent an accidental host since other sympatric anurans (i.e., Bufo spp. and G. olivacea) have been reported to have a higher prevalence of C. variabilis (McAllister and Upton, 1987a; McAllister et al., 1989), whereas the more closely related P. s. streckeri has not been found to be infected (McAllister, 1987).

Larval intradermal mites, Hannemania sp. Oudemans, 1911, infested 8 (57%) P. clarkii from Dallas, 1 (8%) from Hood, and 2 (15%) from Somervell counties. It is not known why there is such a great disparity in prevalence among the 3 localities. Unengorged or partially engorged larvae were encapsulated by host dermal connective tissue. The majority of capsules was found on the undersides of legs, on the venters, and near the cloacal openings of frogs. Because only larvae were found, it was not possible to determine specific identity. However, H. multifemorala Loomis, 1956, has been reported previously from neighboring Erath County on Great Plains narrowmouth toads, Gastrophryne olivacea, H. dunni Sambon, 1928, is known from eastern Texas on dusky salamanders, Desmognathus auricularus and G. olivacea, and H. eltoni (syn. H. penetrans Ewing, 1931) has been reported from Bexar County on southern leopard frogs, Rana sphenocephala (Loomis, 1956). In addition, Kuntz and Self (1944) reported H. eltoni on G. olivacea from Comanche County, Oklahoma, and Duszynski and Jones (1973) reported 9 species of frogs and toads from New Mexico harboring Hannemania sp.

In conclusion, most of the protozoan parasites of *P. clarkii* are shared with sympatric *P. s.* streckeri, while neither the helminth nor arthropod parasites are shared. Furthermore, a common helminth of the region in other anurans (*Cylindrotaenia americana*) is reported to have a low prevalence in *P. clarkii* but apparently is not harbored by *P. s. streckeri*. An ecological explanation for these comparative data is elusive at present; however, it is similar to that reported by Upton and McAllister (1988) who noted that several *Eimeria* spp. infect *P. s. streckeri* but not *P. clarkii* from the same aquatic environment.

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